

# Ecological and evolutionary impacts of changing climatic variability

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## 20 ABSTRACT

While average temperature is likely to increase in most locations on Earth, many places will simultaneously experience higher variability in temperature, precipitation, and other climate variables. Although ecologists and evolutionary biologists widely recognize the potential impacts of changes in average climatic conditions, relatively little attention has been paid to the potential

25 impacts of changes in climatic variability and extremes. We review the evidence on the impacts of  
increased climatic variability and extremes on physiological, ecological and evolutionary processes  
at multiple levels of biological organization, from individuals to populations and communities. Our  
review indicates that climatic variability can have profound influences on biological processes at  
multiple scales of organization. Responses to increased climatic variability and extremes are likely  
30 to be complex and cannot always be generalized, although our conceptual and methodological  
toolboxes allow us to make informed predictions about the likely consequences of such climatic  
changes. We conclude that climatic variability represents an important component of climate that  
deserves further attention.

35 *Key words:* climate change, community structure, demography, fitness, geographic range limits,  
phenological mismatches, phenotypic plasticity, physiological traits, population dynamics, species  
interactions.

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## I. INTRODUCTION

The Earth's climate is changing rapidly. Available data indicate a trend towards increasing global temperatures. Causes for these temperature changes are intricate and associated with a network of

events in which human activities appear to play a determinant role (Trenberth, 2011). In addition to  
45 increased temperature averages, human-caused climate change is expected to lead to increased  
climatic temporal variability and increased occurrence of climatic extremes in many places (e.g.  
droughts, excessive precipitation, heat waves; Easterling *et al.*, 2000; Burroughs, 2007; Meehl *et al.*,  
*et al.*, 2007; Coumou & Rahmstorf, 2012). Under some climate change scenarios, climatic variability  
and extremes are expected to increase more than climatic averages for some regions (Meehl *et al.*,  
50 2007).

A mechanistic understanding of the biological consequences of climate change requires the  
integration of physiology, ecology, and evolutionary biology (Spicer & Gaston, 1999). Although  
biologists widely recognize the potential impacts of changes in average climatic conditions,  
relatively little attention has been paid to the potential impacts of changes in climatic variability and  
55 extremes (Benedetti-Cecchi, 2003; Jentsch, Kreyling & Beierkuhnlein, 2007; Bozinovic, Calosi &  
Spicer, 2011*b*; Thompson *et al.*, 2013), and we know relatively little about the mechanisms  
underlying these potential impacts. Enhancing this understanding and highlighting gaps in our  
knowledge is the main goal of this review.

When discussing the ecological and evolutionary implications of increased climatic  
60 variability and climatic extremes, it is important to define precisely these attributes of climate.  
Climatic variability can be defined as the standard deviation or the variance of a climate variable  
such as temperature, or by a relativized measure of variability such as the coefficient of variation.  
Defining the frequency of occurrence of extreme climatic events in statistically precise terms is  
more elusive. Often extremes are defined in relation to a climate record for a certain period (e.g.  
65 three standard deviations away from the mean temperatures from 1960 to 2010). However, this  
definition is problematic, as it implies that an increase in the mean temperature, with no change in  
the shape of the distribution around the mean, would increase the frequency of high extremes and  
decrease the frequency low extremes, whereas a change in climatic variability might lead to

increased extremes at both ends of the distribution. Alternatively, extremes can be defined as events  
70 that influence the shape of the distribution of a climatic variable without influencing the mean and  
the variance—i.e. the kurtosis (Fig. 1). Thus, an increase in the frequency of extreme events would  
make the distribution of the climatic variable more leptokurtic, while a decrease in the frequency of  
extreme events would make the distribution more platykurtic, without necessarily changing the  
mean or the variance of the climatic variable.

75 Here we review the evidence on the impacts of increased climatic variability and extremes  
on physiological, ecological and evolutionary processes at multiple levels of biological  
organization, from individuals to populations and communities. Specifically, we consider the  
responses to increased climatic variability and extremes in terms of (i) physiology and performance  
including Darwinian fitness, (ii) phenotypic plasticity, (iii) demography and population dynamics,  
80 and (iv) community structure and dynamics.

## II. PHYSIOLOGY, INDIVIDUAL PERFORMANCE AND FITNESS

### (1) Individual performance and fitness

Recent models of the physiological, ecological and evolutionary responses of organisms to  
85 climate change indicate that a change in thermal variance could have as much (or more) of an  
impact on fitness as does a change in the mean temperature (Bozinovic *et al.*, 2013a; Bozinovic,  
Catalán & Kalergis, 2013b; Estay, Lima & Bozinovic, 2014). The typically unimodal, asymmetric  
shape of the relationship between temperature and performance, which tends to accelerate below the  
optimum temperature but decelerate above the optimum, implies that the impact of thermal  
90 variation on physiological performance will depend on the mean temperature in relation to the  
temperature at which performance peaks (Bozinovic *et al.*, 2011a). When mean temperature is  
below the thermal optimum, thermal variation can either enhance or reduce physiological  
performance; by contrast, when mean temperature is equal to or greater than the thermal optimum,

thermal variation will always impair performance (Fig. 2). Thus, the mean and variance of  
95 temperature will interact predictably to determine Darwinian fitness in a variable environment.

Several studies provide experimental evidence for the influence of thermal variance on individual performance. Bozinovic *et al.* (2011a) observed that thermal variation enhanced the population growth rate of the fruit fly *Drosophila melanogaster* at a low mean temperature, while it decreased this rate at the thermal optimum, despite the fact that flies improved their heat and cold  
100 tolerances through environmental acclimation. Siddiqui, Barlow & Randolph (1973) reported significantly different effects of constant and alternating temperatures on population growth of the pea aphid *Acyrtosiphon pisum*; in addition, the net reproductive rate ( $R_0$ ) and the intrinsic rate of increase ( $r$ ) differed in their response to different temperature regimes. More recently, Clavijo-Baquet *et al.* (2014) found similar results for the effects of thermal mean and variability on  $R_0$ ,  $r$  and  
105 the generation length  $T_g$ , of *D. melanogaster*, with no effects of temperature variation on  $r$  despite strong effects on  $R_0$  and  $T_g$ . Thus, considering that  $r \approx \ln(R_0)/T_g$ , thermal effects on  $R_0$  and  $T_g$  cancel each other. Likewise, Kjaersgaard *et al.* (2013) reported that increased temperature variability decreased the individual performance of yellow dung flies *Scatophaga stercoraria*. Furthermore, some studies have shown that moderate amounts of environmental variability do not  
110 necessarily have detrimental effects on performance, compared to greater levels of variability (Pétavy *et al.*, 2004; Folguera *et al.*, 2011). Although experimental approaches cannot capture the wide range of abiotic and biotic conditions in nature, they help us to understand the mechanisms by which climate change may impact organisms. This kind of experiment can also shed light on geographic variation in phenotypes, because diel and seasonal patterns of environmental  
115 temperature change along latitudinal and altitudinal gradients (Angilletta, 2009). Indeed, Kjaersgaard *et al.* (2013) advocate the use of more relevant fluctuating temperatures in experimental studies of the ecological impact of climate change.

It is important to note that individual performance can change with increased temperature

variability even if average temperature does not change. This is because of Jensen's inequality, a  
 120 property of nonlinear functions, which states that, for a sample of  $x$  values with mean  $\bar{x}$  and  
 a nonlinear function  $f(x)$ ,  $\overline{f(x)} > f(\bar{x})$  if  $f(x)$  is accelerating (second derivative is positive)  
 and  $\overline{f(x)} < f(\bar{x})$  if  $f(x)$  is decelerating (second derivative is negative; Ruel & Ayres, 1999).  
 Thus, the typically unimodal, asymmetric shape of the relationship between body temperature and  
 physiological performance (Fig. 2) should lead to complex effects of thermal variance (Estay *et al.*,  
 125 2011): close to the peak of the function, greater temperature variability should decrease  
 physiological performance, whereas at the lower tails of the function greater temperature variability  
 should increase performance. For example, Siddiqui & Barlow (1972), conducted a now classic  
 study that showed that thermal fluctuations, within the range of temperatures favourable for  
 reproduction, enhanced the rate of population growth. Later, Orcutt & Porter (1983) observed a  
 130 slightly positive effect of thermal variation on the population growth rate of water fleas *Daphnia* sp.  
 This result could be attributed to the range of experimental temperatures used, which was in the  
 accelerating (left tail) part of the performance curve. In turn, Estay *et al.* (2011) found that thermal  
 variation had a negative effect on the population growth of flour beetles *Tribolium confusum* when  
 mean temperature was in the decelerating part of the performance curve. Similar interactions  
 135 between the mean and the variance of temperature have been observed in other studies (Dallwitz,  
 1984; Paaijmans *et al.*, 2010; Bozinovic *et al.*, 2013a; Estay *et al.*, 2014).

A mechanistic understanding of organismal responses to climatic variability requires  
 elucidating the underlying cellular and physiological processes that allow animals to cope with such  
 variation. When exposed to stress, organisms respond in multiple ways, including DNA and protein  
 140 repair, apoptosis, removal of cellular and molecular waste generated by stress, and changes in  
 cellular metabolism to switch from cellular growth to repair (Kassahn *et al.*, 2009). Thus, organisms  
 are constrained by a trade-off between response to stress and allocation to growth and reproduction  
 (Somero, 1995). A key component of such responses are heat shock proteins, which have an

important function in cells under stressful conditions and are necessary for the survival and  
145 recovery of organisms, often by rescuing critical metabolic enzymes from destruction (Parsell &  
Lindquist, 1994). Experiments have shown that small amounts of induced heat shock proteins can  
influence development, life span, fecundity and stress resistance (Sørensen *et al.*, 1999). Therefore,  
the influence of heat shock proteins has the potential to scale up to populations and communities.  
The very existence of these molecular and integrative responses to climate suggests that thermal  
150 fluctuation may be an important selective factor in nature. In addition, the effects of different  
thermal conditions are directly related to the thermal safety margin—i.e. the difference between an  
organism's thermal optimum and its current environmental temperature (Deutsch *et al.*, 2008;  
Folguera, Bastías & Bozinovic, 2009). Future interpopulation comparisons might reveal higher  
sensitivity to thermal amplitude in, for instance, lowland than in highland populations, indicating  
155 that important effects on biodiversity may be expected in the context of increasing thermal  
amplitude.

## **(2) Macrophysiological patterns**

Macrophysiology seeks to explain how physiological traits are affected by environmental variation  
160 over large geographic scales. For instance, compared to tropical taxa, species from temperate  
latitudes are thought to experience selection for greater plasticity because they live in more seasonal  
environments (Chown, Gaston & Robinson, 2004; Ghalambor *et al.*, 2006). The latter is the case  
particularly in northern latitudes, where differences between absolute maximum and absolute  
minimum temperatures are greatest (Addo-Bediako, Chown & Gaston, 2000; Chown *et al.*, 2004).  
165 Supporting evidence of a hemispheric asymmetry in physiological tolerance (with assumed  
underlying plasticity) has been reported for insects (Addo-Bediako, Chown & Gaston, 2002;  
Sinclair, Addo-Bediako & Chown, 2003) and terrestrial isopods (Bozinovic *et al.*, 2014), but  
evidence for other taxa is lacking.



Data on latitude and thermal amplitude show that the increased temperature variation in temperate regions results from the pattern of variation of minimum temperatures, with maximum temperatures being equally variable across latitude (Chown *et al.*, 2004b; Ghalambor *et al.*, 2006). Therefore, the expected increased selection on plasticity in temperate species—compared to tropical taxa—should occur for responses to cold and not to warming. This prediction has been corroborated for insects, frogs, and lizards (see Ghalambor *et al.*, 2006). In addition, Vasseur *et al.* (2014) recently demonstrated the independent and interactive effects of mean temperature and its variability on thermal performance curves for nearly 40 ectothermic invertebrates with worldwide distributions. With the use of fine-grained, site-specific historical temperature data, Vasseur *et al.* (2014) showed how changes in the mean, variance and positive skewness of historical temperatures have significant influences on organismal performance. Furthermore, for ectotherms, thermal performance—and hence fitness—tends to decline or remain unchanged under future climate scenarios, with greater declines occurring at mid to low latitudes (see also Deutsch *et al.*, 2008).

### III. PHENOTYPIC PLASTICITY

#### (1) Phenotypic plasticity and adaptation to changing environments

Increased climatic variability may be envisioned as a particular case within the general phenomenon of changing environments. Adaptation of organisms to changing environments occurs mainly by two mechanisms: (i) genetic differentiation, i.e. across-generations phenotypic adjustment to the prevailing environmental conditions as a result of selection on heritable traits, and (ii) phenotypic plasticity, i.e. within-generation phenotypic changes induced by the environment (Schlichting & Pigliucci, 1998; Réale *et al.*, 2003). Evidence strongly suggests that phenotypic plasticity, which may even be reversible (Garland & Kelly, 2006), is essential for adaptation to climate change and increased climatic variability, and that evolutionary rates cannot always keep up with climate change (Jump & Peñuelas, 2005; Davis, Shaw & Etterson, 2005; Gienapp *et al.*, 2008; Charmantier

*et al.*, 2008; Ozgul *et al.*, 2010; Hoffmann & Sgrò, 2011; but see Visser, 2008). For instance, the  
195 advancement in parturition date of the North American red squirrel, *Tamiasciurus hudsonicus*, in  
response to climate change is mostly due to phenotypic plasticity (62%), while microevolutionary  
changes account for 13% of the phenotypic change (Berteaux *et al.*, 2004; Anderson *et al.*, 2012). In  
a modelling study, Chevin, Lande & Mace (2010) found that the critical rate of environmental  
change, i.e. the maximum rate of sustained environmental change that allows population  
200 persistence, increases with decreasing absolute difference between phenotypic plasticity and the  
environmental sensitivity of selection (the change in the optimum phenotype with the environment).  
In other words, the closer plastic responses in a given population track the shift in the favoured  
phenotype that occurs when the environment changes, the greater magnitude of sustained  
environmental change this population will tolerate. Nonetheless, it is likely that rapid responses to  
205 environmental change *via* phenotypic plasticity may be followed by selection and microevolution  
(Matesanz, Gianoli & Valladares, 2010; Godoy *et al.*, 2011; Brunet & Larson-Rabin, 2012).

Phenotypic plasticity is expressed in the reaction norm: the repertoire of phenotypic  
responses to environmental variation of a given genotype, population, or species (Pigliucci, 2001;  
Gianoli & Valladares, 2012). The reaction norm itself may evolve by natural selection (Scheiner &  
210 Lyman, 1991; Pigliucci & Schlichting, 1996; Scheiner & Callahan, 1999; Garland & Kelly, 2006;  
Baythavong & Stanton, 2010). Selection on reaction norms and performance curves in a population  
should be driven by the frequency (and predictability) of the environments experienced by  
organisms (Weis & Gorman, 1990; Gilchrist, 1995; Alpert & Simms, 2002; Kingsolver *et al.*, 2007).  
Phenotypic plasticity should be important not only to deal with the increasing rate of climate  
215 change, but also with the increased unpredictability of climatic anomalies (Arias, Poupin & Lardies,  
2011). An experimental study with ten sub-Antarctic weevil species found that phenotypic plasticity  
accounted for most of the between-population variation in critical thermal minimum (CT<sub>min</sub>) and  
critical thermal maximum (CT<sub>max</sub>) (Klok & Chown, 2003). Weevils acclimated for only one week at

15 °C raised their  $CT_{max}$  by 2.4 °C, compared to those maintained at 0 °C (Klok & Chown, 2003).

220 Interestingly, Skelly *et al.* (2007) estimated the potential for evolutionary response in  $CT_{max}$  in ectotherms and predicted an increase in 3.2 °C across 10 generations (50 years). Thus, within a single generation plastic responses drive an increase in  $CT_{max}$  that is 75% of that calculated for 10 generations when only evolutionary responses are considered. Furthermore, an extreme environment may allow the expression of formerly ‘hidden’ parts of the reaction norm, which may  
225 reveal the existence of cryptic genetic variation upon which natural selection may act (Schlichting, 2008). Overall, in view of the problem of decreased performance in ectotherms caused by climatic variation illustrated in Fig. 2, adaptive phenotypic plasticity may be conceived to function as a buffer to minimize the coupling of environmental temperature and body temperature, in a context of increased climatic variability.

230

## **(2) Patterns of phenotypic plasticity at the population level**

Theory predicts that increased environmental variation should select for increased phenotypic plasticity (Schlichting & Pigliucci, 1998; Alpert & Simms, 2002). Some studies have proved the increased magnitude and adaptive value of phenotypic plasticity in spatially heterogeneous  
235 environments (Lind & Johansson, 2007; Hollander, 2008; Baythavong, 2011), but in the context of climatic variation associated with climate change it is more relevant to focus on temporal heterogeneity. We surveyed published studies to evaluate this theoretical prediction in plant populations from habitats with contrasting temporal environmental heterogeneity. Plants are appropriate study subjects to evaluate this prediction because they are sessile, which means they  
240 cannot migrate to avoid harsh climatic conditions (Post *et al.*, 1999; Réale *et al.*, 2003; Battisti *et al.*, 2006). We specifically focused on those studies reporting plant responses to temperature and soil moisture, which are the components of climate change that are more relevant to plants (Matesanz *et al.*, 2010). We found 11 studies that met these conditions, including 14 cases from 12

species (Table 1). Phenotypic plasticity data included physiological, morphological, and life-history  
245 traits (references in Table 1).

Results support the theoretical expectation that increased plasticity is selected for in more  
variable environments. There was a significant positive association between phenotypic plasticity in  
plant populations and temporal heterogeneity in rainfall or temperature (10 out of 14 cases; Table  
1). These preliminary results with plant species suggest that increased phenotypic plasticity may  
250 drive adaptation to increased climatic variability in natural populations. A broader sample of studies  
is needed to determine whether other taxa respond similarly, and whether the detection of this  
relationship is influenced by the type of traits studied or the intrinsic characteristics of the  
organisms.

### 255 **(3) Evolution of increased plasticity**

How could increased climatic variability select for increased phenotypic plasticity? Schlichting &  
Pigliucci (1998) discuss two general models for genetic control of plastic responses: allelic  
sensitivity and gene regulation. Allelic sensitivity refers to changes in the amount of a given gene  
product in response to changes in the environment, and is considered a 'passive' form of plastic  
260 response that is not necessarily adaptive (Schlichting & Pigliucci, 1998). This mechanism has been  
found in insects (Schmalhausen, 1949), bacteria (Hartl & Dykhuizen, 1981), plants (Kliebenstein,  
Figuth & Mitchell-Olds, 2002) and nematodes (Gutteling *et al.*, 2007), and is expected to be  
involved in plastic responses to temperature because of the close link between enzyme activity and  
temperature (Schlichting & Pigliucci, 1998). By contrast, gene regulation involves the existence of  
265 an environment-dependent regulatory switch in gene expression, resulting in a threshold response,  
and is considered an 'active' form of plastic response, most likely of adaptive nature (Schlichting &  
Pigliucci, 1998). Examples of taxa in which this mechanism has been observed include plants  
(Doebley, Stec & Gustus, 1995), insects (Fairbairn & Yadlowski, 1997), molluscs (Egg *et al.*, 2009),

and bacteria (Rivera-Gómez, Segovia & Pérez-Rueda, 2011). These two mechanisms are not  
270 mutually exclusive, as shown by Wu (1998) for growth traits in poplar trees. Furthermore,  
epigenetic phenomena, i.e. heritable changes in gene expression that are not mediated by changes in  
DNA sequence (Richards, 2006), may mediate functional responses to the environment (Bossdorf,  
Richards & Pigliucci, 2008; Chinnusamy & Zhu, 2009; Bräutigam *et al.*, 2013; Herman *et al.*,  
2014). Examples of epigenetic modifications driving phenotypic plasticity include flowering  
275 regulation in response to cold exposure in *Arabidopsis thaliana* (Kim & Sung, 2012), biomass  
allocation responses to water shortage in poplars (Gourcilleau *et al.*, 2010), niche width in a flower-  
living yeast (Herrera, Pozo & Bazaga, 2012), and transgenerational induction of plant defences  
against herbivores and pathogens (Holeski, Jander & Agrawal, 2012).

Gene regulation should be particularly advantageous for non-instantaneous plastic  
280 responses, such as those involving biomass allocation, whose complete expression involves a time  
lag between the appearance of the environmental cue (the stimulus) and the deployment of the  
target phenotype in the selective environment. Several studies indicate that reliable cues triggering  
plastic responses and somewhat predictable environments are required for plasticity to be adaptive  
(DeWitt, Sih & Wilson, 1998; Langerhans & DeWitt, 2002; Alpert & Simms, 2002; Van Kleunen &  
285 Fischer, 2005; Ghalambor *et al.*, 2007; Reed *et al.*, 2010; Scheiner & Holt, 2012; van den Heuvel *et al.*,  
*et al.*, 2013). Pigliucci (2001) provides a detailed analysis of evolutionary outcomes concerning  
phenotypic plasticity in the presence and absence of environmental cues, considering the duration of  
temporal fluctuation as compared to generation time. In some masting plant species (those that  
show sporadic reproductive outbreaks that satiate herbivores) synchronous fruiting was associated  
290 with anomalously high temperatures in the summer before seedfall, a cue linked with the La Niña  
climatic phenomenon (Schauber *et al.*, 2002). If the frequency of such anomalously high  
temperatures increases, masting might no longer be adaptive in terms of realized herbivore satiation  
(McKone, Kelly & Lee, 1998). In fact, in most of the reported cases of successful tracking of

climate change, including species from diverse phyla, the climatic shift has been rather monotonic  
295 (Parmesan & Yohe, 2003; Root *et al.*, 2003; Parmesan, 2006; Cleland *et al.*, 2007; Walther *et al.*,  
2007; Charmantier *et al.*, 2008). In general, if in order to adapt to seasonal changes species rely on  
photoperiod alone (e.g. Lambrechts & Perret, 2000), they are likely to fail to adjust to the actual  
environment because of the recent shift (or even uncoupling) in the relationship between  
temperature and photoperiod driven by climate change (Visser, Both & Lambrechts, 2004).  
300 Alternatively, if species rely on temperature alone (e.g. Condon, Chenoweth & Wilson, 2010),  
anomalous high-temperature periods may induce maladaptive responses (Powell & Logan, 2005),  
depending on the threshold of response. A mixed control on plasticity of phenology, by both  
photoperiod and temperature, could be a better strategy; this has been shown for several moth  
species at high latitudes (Valtonen *et al.*, 2011). The interaction between resource availability and  
305 climate as environmental cues is also a potentially advantageous alternative to track climate change-  
driven anomalies better (Visser *et al.*, 2004).

Returning to the question of how increased climatic variability could select for phenotypic  
plasticity, and assuming a gene regulatory system underlying the plastic responses, we show in a  
conceptual model (Fig. 3) the likely outcomes for two climate change scenarios: (i) a temporal shift  
310 in environmental quality, and in its corresponding cue for the initiation of the plastic response (Fig.  
3A), and (ii) a temporal shift in environmental quality plus climatic variability (a short-term  
anomaly counteracting the climatic trend; Fig. 3B). Our analysis considers three genotypes with  
different phenotypic responses to the environment across the season (Fig. 3C,D). The black  
genotype is not responsive to climate change or climatic variability, thus maintaining the phenotypic  
315 trajectory that matches the historical climate. The red genotype tracks both climate change and  
climatic variability, adjusting its reaction norm accordingly. The purple genotype adjusts its reaction  
norm, as for the red genotype, and shows increased plasticity, envisioned as a likely response to an  
increasingly heterogeneous environment. We then estimate the relative advantage of each strategy

(genotype) considering two possible adaptive landscapes: directional and stabilizing selection  
320 (Endler, 1986). When directional selection prevails, increased phenotypic plasticity (purple) is  
favoured under both climatic scenarios, especially under climatic variability (Fig. 3E,F). By  
contrast, if stabilizing selection prevails, increased plasticity is not beneficial, being as  
(mal)adaptive as the unresponsive black genotype under both climatic scenarios; the red genotype is  
favoured instead. Therefore, whether increased plasticity is favoured would ultimately depend on  
325 the form of natural selection acting at the end of the season. The latter is influenced by both the  
selective factors operating and the traits involved (Endler, 1986; Kingsolver *et al.*, 2001; Geber &  
Griffen, 2003), which may show ontogenetic variation in their environmental sensitivity (e.g. Arias  
*et al.*, 2011). Thus, temperature-related performance traits regularly show an optimum, with low and  
high levels having a lower adaptive value (Huey & Kingsolver, 1989; Angilletta, Niewiarowski &  
330 Navas, 2002; Seebacher, 2005); by contrast, water-economy traits often show linear relationships  
with performance (Dudley, 1996; Heschel *et al.*, 2004; Saldaña *et al.*, 2007). Nonetheless,  
stabilizing selection is less frequent and usually weaker than directional selection in natural  
populations (Kingsolver *et al.*, 2001). In any event, increased levels of climatic complexity (the  
particular sequence and duration of environmental states) may complicate patterns of selection on  
335 thermal performance curves and other continuous reaction norms (Valladares, Gianoli & Gómez,  
2007; Kingsolver *et al.*, 2007). Moreover, a comprehensive and realistic analysis must take into  
account the role of correlations between characters of functional value, which may constrain the  
evolution of adaptive responses to climate change (Etterson & Shaw, 2001; Gianoli & Palacio-  
López, 2009).

340 The above analysis may allow identifying the different targets involved in adaptive  
evolution of phenotypic plasticity to deal with climatic variability. First, it may entail the evolution  
of responsiveness (the red genotype), i.e. a shift in the environmental threshold of response or a  
change in the eliciting cue. This has been shown for the flowering time in an annual plant, with

summer drought selecting for early flowering and causing adaptive evolution in a few generations  
 345 (Franks, Sim & Weis, 2007). Second, it may involve the evolution of increased plasticity (the purple  
 genotype), i.e. a reaction norm with a steeper slope. This has been demonstrated for an  
 insectivorous bird, which was able to change the duration of components of breeding behaviour in  
 order to track the temporal shift in food availability (Charmantier *et al.*, 2008). Finally, adaptation  
 to climatic variability may include the arrestment or reversion of phenotypic changes, once the  
 350 climatic anomaly is detected (the red and purple genotypes). Of course, this is possible only for  
 certain traits, with behaviour appearing as a suitable candidate. Besides the above-mentioned study  
 on adaptive responses to climate change in bird breeding behaviour (Charmantier *et al.*, 2008), it  
 has been reported for the same species that (i) there is no response of birds to warming in  
 physiological or morphological traits (Visser *et al.*, 2011), (ii) the cue is increased temperature and  
 355 not mean temperature (Schaper *et al.*, 2012), and (iii) there is genetic variation for cue sensitivity  
 (Visser *et al.*, 2011), thus providing the basis for an evolutionary response to natural selection. In a  
 related species, Vedder (2012) recently showed experimentally that birds are able to advance  
 offspring hatching in response to increased temperature after the start of laying. Can other groups  
 do equally well when the trait is other than behaviour? Further research is needed to address this  
 360 question (see Parmesan, 2006; Skelly *et al.*, 2007; Visser, 2008; Hoffmann & Sgrò, 2011).

#### IV. DEMOGRAPHY AND POPULATION DYNAMICS

##### (1) Review of relevant theory

Population theory has primarily examined how changes in environmental variability will influence  
 365 the average rate of annual population growth over the long term, the so-called stochastic population  
 growth rate,  $\lambda_s$  (Caswell, 2001). This rate represents the appropriate average multiple by which  
 the size of the population changes from one year to the next over a long period of time during which  
 the actual population growth rates differ from year to year because of environmental variation. If



$\lambda_s > 1$  , the population tends to increase over the long term, while if it is below one, the population  
 370 tends to decline. Year-to-year variation in the population growth rate arises because of variation in  
 the ‘vital rates’ (survival, somatic growth, and reproduction of individuals in all life stages, plus  
 recruitment of new individuals) that collectively govern the loss of existing individuals and the  
 addition of new individuals to the population. Temporal variation in the vital rates is in turn driven  
 by year-to-year variation in environmental conditions (e.g. Doak & Morris, 2010).

375 Ideally, we would like to be able to say whether an increase in the variance of the vital rates  
 driven by changes in environmental variability (keeping the mean vital rates constant) would either  
 increase or decrease  $\lambda_s$  . Unfortunately, population theory demonstrates that a categorical  
 statement about the effects of increasing environmental variability on the population growth rate is  
 impossible to make. Instead, the answer is that increasing variability can either increase or decrease  
 380 the rate of population growth over the long term, depending on numerous details about how the  
 environment varies and about the demography of the species in question (Fig. 4). We now briefly  
 summarize the most important of these details.

The first thing we need to know to predict if increasing environmental variability would  
 increase or decrease the long-term population growth rate is how the environment varies. The  
 385 simplest case is that environmental variability is independently and identically distributed (IID),  
 meaning that environmental conditions (or more precisely the resulting vital rates) are drawn each  
 year from a statistical distribution with a constant (i.e. ‘identical’) shape and in which the value in  
 one year is drawn ‘independently’ from the values in all previous years. We would then be asking  
 how a step-like increase in the variance of the distribution (i.e. a one-time change in the variance to  
 390 a new, higher value) affects  $\lambda_s$  . In the simplest case in which the population is ‘unstructured’ (i.e.  
 all individuals contribute equally to population growth) and generations do not overlap, increasing  
 the variance of the annual population growth rate (i.e. the degree to which births exceed deaths)  
 decreases  $\lambda_s$  , because  $\lambda_s$  is the geometric mean of the annual population growth rates, and the

geometric mean of any set of numbers decreases as the variance of those numbers increases  
395 (Lewontin & Cohen, 1969). This argument, which ignores the complexity of demography in a  
structured population, would suggest that increasing environmental variability is detrimental to  
long-term population growth.

However, in the case of a structured population, even in the simple and often unrealistic case  
of IID environmental variability, increased variance of a single vital rate can either increase or  
400 decrease  $\lambda_s$ , depending on two demographic factors. First, it depends on whether that vital rate  
covaries, either positively or negatively, with other vital rates. Positive covariation means that the  
two rates tend to be relatively high in the same years (and both relatively low in the other years),  
and negative covariation means that one vital rate tends to be low when the other is high (and *vice*  
*versa*). A number of biological factors can determine whether vital rates covary, and if so, whether  
405 the covariation is positive or negative. For example, vital rates that represent the same type of  
demographic process (e.g. survival of newborns and survival of adults) might respond similarly to  
environmental conditions, creating positive covariation between those rates. Negative covariation  
may arise when rates represent opposing types of demographic processes (e.g. years of high  
individual growth are also years of low shrinkage to smaller size in plants), or it may reflect life  
410 history trade-offs (e.g. high investment in reproduction leads to low growth in the same year). The  
second demographic factor that determines how  $\lambda_s$  will respond to an increase in vital rate  
variance are the ‘sensitivities’ of the deterministic population growth rate to changes in that vital  
rate and in all other vital rates with which it covaries (de Kroon *et al.*, 1986; Caswell, 2001).  
Graphically, a sensitivity represents the slope of a graph of the population growth rate versus the  
415 value of a vital rate, with all vital rates at their current mean values (see Chapter 9 in Morris &  
Doak, 2002). The sensitivities play an important role, because they govern the degree to which  
year-to-year variability in the vital rates translates into variability in the annual population growth  
rate, which in turn influences  $\lambda_s$ . Specifically, variation in vital rates with high-magnitude

sensitivity produces greater variation in the population growth rate. The sign of the sensitivity also  
 420 indicates whether increasing a vital rate increases (as for individual growth rate) or decreases (as for  
 shrinkage) the population growth rate. When two vital rates covary, we must know if the signs of  
 their sensitivities are the same or different to predict the effect of increasing the variance of one of  
 those rates. But if we know the environment is IID, and have estimated the means, variances,  
 covariances, and sensitivities of all the vital rates, we can predict whether increased variance in a  
 425 vital rate will increase or decrease  $\lambda_s$ , as laid out in Fig. 4. These results rely on Tuljapurkar's  
 (1990) 'small-noise' approximation for  $\lambda_s$ , as explained in greater detail in Appendix A1 (see  
 Section IX).

However, environmental variation may not be IID. One common case in which it is not is  
 represented by ecosystems influenced by repeated cycles of disturbance (such as fires or hurricanes)  
 430 and recovery. Here, environmental conditions, and therefore vital rates, are correlated across years  
 because, following a disturbance, recruitment opportunities and individual growth tend to be high  
 for several years running, while late in the disturbance cycle when competition is intense,  
 recruitment and growth are both persistently low (e.g. Menges & Quintana-Ascencio, 2004; Menges  
*et al.*, 2006). In such Markovian environments (so-called because we use a Markov matrix to  
 435 govern the choice of next year's environmental state—for example, the number of years since the  
 last fire—given the environmental state this year, which may influence the probability of another  
 fire), variation even in high-sensitivity vital rates can be adaptive, and thus greater—not less—  
 variation can lead to higher population growth (Tuljapurkar, Horvitz & Pascarella, 2003; Haridas &  
 Tuljapurkar, 2005). For example, in a fire-prone ecosystem, high survival of seedlings immediately  
 440 after a fire followed by low seedling survival once full vegetation cover has been restored and  
 opportunities for recruitment have vanished may yield a higher long-run population growth rate  
 than would the same average rate of seedling survival every year, regardless of the stage in the fire  
 cycle. If so, climate change leading to less variation in survival among the phases of the fire cycle

could be detrimental to population growth, as Morris *et al.* (2006) predicted for two species of fire-  
445 adapted plants. But to add further complication, greater environmental variation at *each* stage of the  
disturbance/recovery cycle (e.g. due to an increase in precipitation variability unrelated to fires)  
could reduce population growth even while greater variation *between* disturbance phases increases  
population growth. Morris *et al.* (2006) also observed this conflicting effect of increasing the two  
kinds of variability in the two study plants, although the effect of changing between-phase  
450 variability was greater, suggesting that reducing both kinds of variability simultaneously would  
depress population growth.

An important caveat is that the results in the preceding paragraph assume no change in the  
frequency of disturbance; they apply only to changes in the between- and within-disturbance phase  
variabilities of the vital rates at a fixed disturbance frequency. Yet changes in climatic variability  
455 will often drive changes in the frequency of disturbance (e.g. making fires or hurricanes more or  
less likely). When the disturbance frequency changes, it becomes meaningless to ask how changing  
vital rate variability *per se* will affect  $\lambda_s$ . The reason is that changing the disturbance frequency  
will simultaneously change the means and the variances of the vital rates, so we can no longer  
assess the effect of changing the variance while holding the mean constant, as we have done so far.  
460 Changing disturbance frequency will also change the temporal correlations within and between vital  
rates (these correlations measure how similar a vital rate's values are from one year to the next, as  
well as the similarity in one vital rate in one year to all the other vital rates in the next year). All of  
these changes to means, variances, and temporal correlations will collectively result in changes to  
 $\lambda_s$  (Tuljapurkar & Haridas, 2006).

465 Given this complexity, it is possible for changing disturbance frequency to change the  
variance of a vital rate and  $\lambda_s$  in the same or in opposite directions. For example, consider the  
situation in which conditions are best for a species immediately after a disturbance, as would likely  
be the case for a fugitive species. Increasing the disturbance frequency would then be expected to

increase the long-term population growth rate, but decrease the vital rate variances (e.g. Caswell & Kaye, 2001; Tuljapurkar *et al.*, 2003; Horvitz, Tuljapurkar & Pascarella, 2005; Tuljapurkar & Haridas, 2006). Other species may show poorer population performance immediately after a disturbance but better performance at intermediate times since the last disturbance. For example, a fire may kill aboveground plants but spur recruitment from the seed bank. If recruits take several years to reach reproductive size, too high a fire frequency would preclude reproduction and prevent replenishment of the seed bank. In this scenario, an increase in disturbance frequency could decrease both  $\lambda_s$  and the vital rate variances (e.g. see Gross *et al.*, 1998; Satterthwaite, Menges & Quintana-Ascencio, 2002; Menges & Quintana-Ascencio, 2004; Menges *et al.*, 2006). In general, the variance of a vital rate as a function of the disturbance frequency should follow a unimodal curve, being low when disturbance frequency is low (and environmental conditions are almost always in the undisturbed state) or when it is high (and the environment is nearly always disturbed), but highest at an intermediate disturbance frequency (when disturbed and undisturbed environmental states alternate frequently). Whether increasing disturbance frequency will increase or decrease  $\lambda_s$  and the vital rate variances will depend on the current disturbance frequency as well as whether the population performs best at low, high, or intermediate disturbance frequency.

Finally, environmental variability may be neither IID nor Markovian. One important example is temporal trends in the mean and/or the variance of a climatic driver of demography, resulting in shifts in the means and/or variances of vital rates. Importantly, the mean and variance of a vital rate need not both increase or both decrease, but may change in opposite directions (Fig. 5). When the distribution from which annual environmental conditions are drawn is continuously shifting, the results summarized above may only serve as a general guide to the short-term response of the population growth rate, and computer simulations with changing means and/or variances would be required to predict the effective population growth rate—or population size—over time (Fig. 4).

To summarize what population theory has to say about the likely effect of changing  
495 environmental variability, greater variability can either increase or decrease the long-term rate of  
population growth. To predict the outcome, in addition to characteristics of the environmental  
variability and of the demography of the species in question, we may also need to consider  
simultaneous changes in the means and temporal correlations of environmental drivers, especially  
in the face of changing disturbance frequency or of long-term trends in the distribution of  
500 environmental conditions.

## **(2) Empirical estimates of population sensitivity to increased environmental variability**

Another way to assess the potential effects of increased climatic variability on population growth is  
to compute the so-called stochastic sensitivities of the long-run population growth rate to the  
505 standard deviations of the vital rates (Tuljapurkar *et al.*, 2003). These are different from the  
sensitivities of the deterministic population growth rate to the vital rate means that were described  
in the preceding section; they represent the slope of the long-run stochastic growth rate with respect  
to the temporal standard deviation of a vital rate, evaluated at the current standard deviation. The  
computation of the stochastic elasticities (Tuljapurkar *et al.*, 2003) can account for any temporal  
510 correlations in environmental conditions, and therefore vital rates, in a Markovian—indeed in any  
auto-correlated—environment. Using multi-year demographic data, Morris *et al.* (2008) computed  
the stochastic sensitivities to changes in the means and standard deviations of vital rates for 36  
animal and plant species ranging from insects to mammals and annual to long-lived perennial  
plants. In general, the population growth rates of these species are more sensitive to climate-driven  
515 changes in the vital rate means than they are to changes in vital rate standard deviations driven by  
changes in climatic variability. Nevertheless, short-lived species showed relatively greater  
sensitivity to increasing vital rate variability than did long-lived species. This result suggests that an  
increase in climate variability might have a disproportionately negative impact on the demography

of short-lived species. Conversely, in regions expected to experience reduced climatic variability,  
520 problems for humans and other species posed by (mostly short-lived) agricultural pest insects and  
disease vectors, and by invasive annual plants, may worsen.

It is important to emphasize that the preceding paragraph describes what *could* happen to the  
study populations should climatic variability change. We still have no definitive examples in which  
a change in the temporal variability of vital rates has been demonstrated to have resulted from a  
525 change in climatic variability. Showing that variability in vital rates has changed is substantially  
more difficult than documenting a trend in the mean of a vital rate over time, because we need a  
series of years of demographic data just to make one reliable estimate of the variability of a vital  
rate, and at least two such series to assess whether variability has changed. But as the number of  
long-term demographic studies has increased in recent years (Crone *et al.*, 2011), we may  
530 eventually be able to correlate changes in vital rate variability with changes in climate variability. In  
the shorter term, if we can correlate particular vital rates with specific climatic drivers, then is it  
probably safe to conclude that changes in the variabilities of those drivers will cause the  
variabilities of the vital rates to change, with the likely population consequences reviewed above.

A less direct way to assess whether increasing climatic variability will elevate variation in  
535 vital rates is to compare the temporal vital rate variances in central and peripheral populations. A  
species may more often experience climatic conditions that are farther from optimum at the range  
margin compared to the range centre (although another possibility is that conditions are always less  
suitable at the margin, or that climate plays no role in setting a particular range limit). If so, vital  
rate variances *may* be higher in peripheral populations, suggesting that even higher climatic  
540 variability beyond the margin may be contributing to the range limit. While some studies have  
found higher demographic variation in peripheral populations (Angert, 2009; Doak & Morris,  
2010), others have not (Villellas, Morris & García, 2013*b*; Villellas *et al.*, 2013*a*).

### (3) Summary and future directions

545 In summary, we cannot make any blanket statements about whether increasing demographic  
variability driven by increasing variability in climate would be detrimental for the rate of population  
growth (or, conversely, whether decreasing variability would be beneficial). The answer depends on  
such factors as the organism's life history (which influences the sensitivities of the vital rates), the  
precise vital rates whose variabilities are changing, whether those vital rates are correlated with one  
550 another, whether the environmental variability is IID *versus* Markovian (as it would likely be in a  
disturbance-prone environment), and if the latter, whether climate change alters the frequency of  
disturbance, the variation between disturbance phases, or the variation within phases. But with  
sufficient demographic data, the modelling tools, some of which have been summarized here, exist  
to predict the effects of changing variability *per se* on population growth.

555 But changes in vital rate variability alone may not capture all of the effects of climate  
change on population growth. Changes in the frequencies of extreme values of the vital rates driven  
by changes in climatic extremes may affect long-run population growth in ways that are not well  
captured by measures of population growth based on small-noise approximations. Instead, the  
demographic consequences of changes in the frequency of extremes may be better explored by  
560 means of computer simulations, drawing vital rates from the 'normal' range of variation with  
occasional (and perhaps increasingly frequent) extreme values 'thrown in', or by slowly changing  
the kurtosis (as well as the mean and variance) of the probability distribution from which each vital  
rate's values are drawn. A yet more promising approach is to link vital rates to environmental  
drivers, so that climatic extremes will produce vital rate extremes directly. Simulating changes in  
565 climatic means, variabilities, and extremes, and then predicting in turn the resulting vital rates,  
population growth (including density-dependent effects), and population size is perhaps the most  
comprehensive way to assess the likely effects of changes in all aspects of climate for the dynamics  
of populations (Ehrlén & Morris, 2015).



## 570 V. SPECIES INTERACTIONS AND COMMUNITY STRUCTURE

### (1) Phenological mismatches, species interactions and geographic range limits

Climate change will influence species interactions and communities either by directly affecting the abundances of interacting species through demographic effects, or by affecting their phenology, which may disrupt the phenological matching among interacting species. We have already discussed  
575 demographic effects in the previous section. Below we discuss how climate change, and particularly increased climatic variability, can lead to phenological mismatches among interacting species, and how those mismatches can influence geographic range limits.

Climate change should lead to phenological mismatches among interacting species if species respond idiosyncratically to climatic variables. If so, changes in one climatic variable may influence  
580 the phenology of one species but not of its interaction partners (Harrington, Woiwod & Sparks, 1999; Visser & Both, 2005; Parmesan, 2006; Devoto, Zimmermann & Medan, 2007; Memmott *et al.*, 2007; Tylianakis *et al.*, 2008; Hegland *et al.*, 2009; Yang & Rudolf, 2010). For example, a plant could respond to increased average temperature by advancing its flowering period (as has been frequently observed; Cleland *et al.*, 2007; Gordo & Sanz, 2010; CaraDonna, Iler & Inouye, 2014),  
585 while an insect pollinator may respond more strongly to photoperiod (which is determined by latitudinal position, not climate), so that its developmental phenology remains unaffected in spite of increased average temperature; this differential response to climate change would result in a phenological mismatch between these species. There are several examples in the literature of how gradual changes in climatic averages have led to phenological mismatches and concomitant  
590 demographic consequences for the interacting species (Visser *et al.*, 1998; Visser & Both, 2005; Both *et al.*, 2009; Plard *et al.*, 2014). Of course, a partial phenological mismatch between interacting species does not necessarily indicate the influence of climate change, as mismatches are known to have occurred naturally before the onset of human-driven climate change, even for

relatively specialized interactions (Singer & Parmesan, 2010); however, such mismatches are likely  
595 to be exacerbated by climate change, which may in turn exacerbate the demographic vulnerability  
of the interacting species.

The ecological consequences of climate-driven changes in the phenological matching of  
interacting species are likely to be qualitatively different under changes in climatic averages and  
changes in climatic variability and extremes. A change in a climatic average may lead to a slow  
600 adjustment of phenologies over decades or centuries (Parmesan *et al.*, 1999; Easterling *et al.*, 2000;  
Parmesan & Yohe, 2003; Root *et al.*, 2003; Thomas *et al.*, 2004; Pounds *et al.*, 2006), which may  
allow species to adapt to changes in the phenology of their interaction partners to minimize  
mismatches (Harrington *et al.*, 1999; Visser & Both, 2005; Parmesan, 2006; Visser, 2008; Hegland  
*et al.*, 2009). By contrast, increased inter-annual variability and the increased occurrence of climatic  
605 extremes may lead to high temporal variation in the phenological matching of interacting species,  
which may impose conflicting selective pressures on interacting species, preventing adaptation and  
maintaining mismatches in the long run. Furthermore, the magnitude of mismatches generated by  
year to year climatic variation is likely to be substantially greater than that generated by gradual  
changes in average climatic conditions.

610 Mismatches in the phenologies of interacting species generated by climate change should  
influence the limits of geographic ranges if they influence demography, and if such influence is  
stronger at range limits than away from them. Under such circumstances, it is at least possible that  
increased climatic variation could lead to a shrinkage of geographic ranges, even if average climatic  
conditions remained unchanged. A greater influence of phenological mismatches on demography at  
615 range limits may occur through at least two distinct mechanisms likely to operate with particular  
strength at range limits: the greater influence of extreme climatic events, and the decreased  
sampling probability of phenological tails.

First, phenological shifts may expose populations to harsh, extreme climatic events, which

are likely to be particularly frequent at range limits; this greater exposure to extreme climatic events  
620 at range limits may impose strong constraints on resource availability and, therefore, should have  
more detrimental demographic effects (Fig. 6A). For example, earlier flowering because of  
increased temperature may also mean a greater likelihood of flower damage due to early spring  
frosts or snows at the upper altitudinal or latitudinal range limits (Inouye, 2008); thus, the same  
phenological shift in flowering may have greater consequences for an animal feeding on these  
625 flowers at its range limit.

Second, if the abundance of resources, mutualists or antagonists decreases towards range  
limits, the demographic consequences of phenological mismatches should also be particularly  
strong at range limits. This is because the high likelihood of not sampling from the tails of the  
phenological distribution when abundance is low leads to a shortening of the phenological period  
630 (Fig. 6B); this is the same sampling issue that makes dates of first flowering unreliable to study  
phenological shifts (van Strien *et al.*, 2008; Miller-Rushing, Inouye & Primack, 2008). A similar  
effect is expected if climatic conditions decrease resource abundance without shifting phenology.  
Thus, for example, if resources are rarer at range limits, a bad year with lower-than-average  
resource abundance may leave individuals with too few resources, sending populations into decline.

635

## **(2) Community structure and dynamics**

Although many studies have discussed the potential effects of climate change on community  
structure and dynamics (e.g. Memmott *et al.*, 2007; Hegland *et al.*, 2009; Woodward *et al.*, 2010),  
the focus of the vast majority of these studies is on climatic averages. Only recently have a few  
640 studies begun to consider how climatic variability and climatic extremes can influence communities  
(e.g. Knapp *et al.*, 2002; Stireman *et al.*, 2005; Benedetti-Cecchi *et al.*, 2006; Jentsch *et al.*, 2007;  
Post, 2013; Reyer *et al.*, 2013; Thompson *et al.*, 2013). For example, Knapp *et al.* (2002) found that  
increased variability in precipitation leads to increased plant diversity in a temperate North

American grassland, while Benedetti-Cecchi *et al.* (2006) showed that temporal variance mitigates  
645 the impact of mean intensity of stress in climate change in Mediterranean algal and invertebrate  
intertidal assemblages. In turn, Stireman *et al.* (2005) have shown that parasitism by insect  
parasitoids on multi-species assemblages of host caterpillars decreases with increasing inter-annual  
variation in precipitation. As is usual in community ecology (see, e.g. Lawton, 1999), our ability to  
draw generalizations about the potential effects of such climatic changes is likely to be contingent  
650 upon the way the community is organized and the type of ecosystem considered. Yet, the few  
available studies give us some hints as to how increased climatic variability and extremes may  
influence community structure and dynamics.

One way in which environmental variability can influence community structure and  
dynamics is through the storage effect, an ecological mechanism that promotes coexistence among  
655 competing species in variable environments (Chesson, 1994, 2000). Three conditions must be met  
for the storage effect to operate. First, species must have some kind of ‘storage’ ability to persist  
during unfavourable periods, such as a seed bank or diapause. Second, species must have  
idiosyncratic (i.e. species-specific) responses to the environment. Third, the effects of intraspecific  
competition on each species must covary with the effects of environmental conditions, so that  
660 competition is stronger in favourable years. Thus, under the storage effect, greater climatic  
variability should promote coexistence and enhance species diversity. Several studies provide  
evidence that the storage effect may be a common, albeit not universal, mechanism promoting  
community stability and diversity. In a North American temperate grassland, Adler *et al.* (2006)  
found that interannual climatic variability promoted the coexistence of three common grass species.  
665 By contrast, a similar analysis in a temperate sagebrush steppe found only weak effects of climatic  
variability on species coexistence (Adler, HilleRisLambers & Levine, 2009). In experimental  
aquatic microcosms, several studies have shown that species coexistence is favoured by increased  
temperature fluctuation (Eddison & Ollason, 1978; Descamps-Julien & Gonzalez, 2005; Jiang &

Morin, 2007); however, the type of environmental fluctuation appears to determine the type of  
670 mechanism involved, with the storage effect operating under red (positively autocorrelated)  
fluctuations and other mechanisms (particularly relative non-linearity; Chesson, 1994, 2000) under  
white (random) fluctuations (Jiang & Morin, 2007).

The type of interaction dominating the organization of the community is likely to influence  
the community's response to climate change. For example, analysing communities with a simple,  
675 tri-trophic structure, Post (2013) found that the trophic level with the weakest self-regulation  
exhibited the strongest response to climatic variation, which may in turn cause instability of the  
entire community. By contrast, in communities structured mainly through competitive interactions,  
climatic variation may help to prevent instability. A similar result was found by Kuang & Chesson  
(2009) for an annual plant assemblage, in which generalist seed predation shifted plant–plant  
680 interactions from resource competition to apparent competition (i.e. potential competitors  
interacting indirectly through common predators; Holt, 1977), thus weakening the storage effect  
that promotes coexistence among plant species.

The size of the community (i.e. the number of species) may also influence the community's  
response to increased climatic variability. In a simulation study of model communities, Kaneryd *et*  
685 *al.* (2012) found that the likelihood of extinction cascades increased with increasing community  
size, particularly under low correlation in species' responses to environmental fluctuations. These  
results suggest that species-rich communities may be less robust to increasing levels of  
environmental variability than their species-poor counterparts.

The breadth of resource use and the degree of specialization/generalization of interactions  
690 are also likely to influence how community structure and dynamics are influenced by climatic  
variability. In particular, greater variability should affect specialists more strongly, because they  
depend on fewer resources and hence should be more susceptible to environmental variability, all of  
which should favour generalists (MacArthur, 1972; Reyer *et al.*, 2013). In turn, the differential

negative effect of climatic variability on specialists could lead to decreased species richness in the  
695 community (MacArthur, 1972; Vázquez & Stevens, 2004), and to a re-arrangement of the  
interaction structure of the community (e.g. increased connectance—the proportion of potential  
links that actually occur).

Different functional groups are also likely to respond differently to increased climatic  
variation. In an analysis of the responses of North American grassland communities to interannual  
700 variation in precipitation, Cleland *et al.* (2013) concluded that systems dominated by perennial  
plants should be more stable to increasing interannual climatic variation, while systems with many  
rare, annual species should exhibit the greatest temporal variability in species composition in  
response to increased climatic variability, especially increased interannual variability in  
precipitation. In aquatic systems, increased temperature variability has been found to lead to  
705 increased richness of zooplankton in lakes (Shurin *et al.*, 2010), while it led to decreased richness of  
phytoplankton in experimental aquatic microcosms (Burgmer & Hillebrand, 2011). Of course, the  
contrasting results of the above studies might simply reflect methodological differences of the  
studies; but it is also possible that they partly reflect the underlying mechanisms that govern the  
dynamics of different functional groups.

710 In consumer–resource interactions, the relative size of the consumer and the resource  
species might also determine the effects of climatic variability on the outcome of interactions and,  
therefore, species coexistence. Rohr *et al.* (2013) have argued that parasites are typically smaller  
and their metabolisms faster than their hosts, which would allow them to achieve faster  
acclimatization to temperature shifts than their hosts, facilitating their transmission and successful  
715 infection. Several studies provide evidence in favour of this conjecture. For example, increased  
climatic temporal variability appears to increase transmission of a fungal parasite of amphibians  
believed to be partly responsible for the drastic declines of many amphibian species worldwide  
(Rohr & Raffel, 2010; Hamilton *et al.*, 2012; Raffel *et al.*, 2013). Similarly, increased climatic

variability may enhance the transmission of malaria (Zhou *et al.*, 2004; Paaijmans *et al.*, 2010),  
720 dengue (Lambrechts *et al.*, 2011), avian influenza (Liu *et al.*, 2007), and cholera (Rodó *et al.*, 2002).

In addition to the ecological characteristics of the community and the interacting species, the  
community's response is also likely to depend on the magnitude of changes in climatic variability  
and extremes. Based on an extensive review of the literature on dryland ecosystems, McCluney *et al.* (2012) predicted that modest increases in the variability of water availability should lead to  
725 strong bottom-up effects in dryland communities, whereas greater increases in the variability of  
water availability should lead to a dominance of top-down effects, and more non-linear and  
unpredictable community dynamics.

## VI. CONCLUSIONS

- 730 (1) Our review indicates that climatic variability can influence biological processes at multiple  
scales of biological organization, from individuals to populations and communities, and from  
physiological to ecological and evolutionary processes. Although responses to climatic variability  
are usually complex and cannot always be generalized, climatic variability represents an important  
component of climate with potentially profound biological consequences.
- 735 (2) At the level of individual performance, the nonlinear relationship between temperature and  
performance implies that the impact of thermal variation on physiological performance will be  
contingent on the temperature at which performance peaks. Thus, close to the peak of the  
performance function, greater temperature variability should decrease physiological performance,  
whereas at the tails of the function greater temperature variability should increase performance. The  
740 relationships among thermal environments, thermal tolerance and population growth may reflect  
natural selection on developmental acclimation. Many researchers have examined geographic  
variation in thermal tolerance, which indicates that genotypes from higher latitudes tolerate low  
temperatures better and recover from thermal shock more rapidly, which probably provides a fitness

advantage. In addition, the ability to recover from thermal shock covaries with environmental  
 745 conditions, suggesting that temperature acts as a selective agent. Because environmental  
 temperature varies over space and time, organisms are continually challenged to maintain  
 homeostasis. Thus, populations are expected to evolve physiological adaptations to local climatic  
 conditions in heterogeneous environments.

(3) Functional phenotypic adjustments to changing environmental conditions may occur *via*  
 750 adaptive evolution across generations and/or within-generation adaptive phenotypic plasticity.  
 Increased environmental variation should select for increased phenotypic plasticity, a prediction  
 supported by our review of studies on plants. Evolutionary responses to selection on plasticity may  
 include a shift in the environmental threshold of response, a change in the eliciting cue, increased  
 magnitude of plasticity (steeper slope of the reaction norm), and even the arrestment or reversion of  
 755 phenotypic changes. The mechanisms underlying such phenotypic patterns may include allelic  
 sensitivity, gene regulation, and epigenetic control. Our conceptual model suggests that the extent to  
 which increased plasticity is favoured by increased climatic variation would ultimately depend on  
 the type of selection operating. Thus, directional selection would clearly favour increased plasticity,  
 while stabilizing selection may render enhanced plasticity maladaptive. However, directional  
 760 selection often prevails over stabilizing selection in the wild. Therefore, the advantageous nature of  
 increased plasticity in the context of increased climatic variation should be verified in most cases.

(4) At the level of populations, the effects of changing climatic variability on population growth can  
 be predicted with the appropriate modelling tools. Whether the effects of increased climatic  
 variation are positive or negative depends on such factors as the organism's life history, the precise  
 765 vital rates whose variabilities are changing, the degree of correlation among vital rates, the type of  
 environmental variability involved (IID *versus* Markovian), the impact of climate change on  
 disturbance frequency, and the degree of variation between *versus* within phases of the disturbance  
 recovery cycle. Furthermore, changes in the frequencies of extreme values of the vital rates driven



by changes in climatic extremes are also important, as they may affect long-run population growth  
770 in ways that are not well captured by measures of population growth based on ‘small-noise’  
approximations. In this case, an alternative approach is the use of computer simulations, drawing  
vital rates from the ‘normal’ range of variation with occasional (and perhaps increasingly frequent)  
extreme values ‘thrown in’, or by slowly changing the kurtosis (as well as the mean and variance)  
of the probability distribution from which each vital rate’s values are drawn. A yet more promising  
775 approach is to link vital rates to environmental drivers, so that climatic extremes will produce vital  
rate extremes directly. Computer simulations may be the only feasible way to link complex changes  
in climatic means, variabilities, and extremes, as well as biotic interactions and intraspecific density,  
to their effects on vital rates, population growth, and the future abundance and distribution of  
species.

780 (5) At the level of species interactions and communities, climate-driven phenological mismatches  
are a key mechanism underlying the potential effects of increased climatic variability and extremes.  
Increased inter-annual variability and the increased occurrence of climatic extremes may lead to  
high year to year variations in the phenological matching of interacting species. The magnitude of  
such mismatches is likely to be substantially greater than that generated by gradual changes in  
785 average climatic conditions, which may impose conflicting selective pressures on interacting  
species, preventing adaptation and maintaining mismatches in the long run. These climate-driven  
phenological mismatches are likely to be especially strong towards the limits of geographic ranges;  
thus, it is possible that increased climatic variation could lead to a shrinkage of geographic ranges,  
even if average climatic conditions remain unchanged. Our ability to draw generalizations about the  
790 potential effects of increased climatic variability at the level of entire communities is rather limited,  
given their inherent complexity; the response of communities to increased climatic variability will  
be most likely influenced by the mechanisms regulating coexistence among interacting species, the  
type of interaction dominating the organization of the community, the community's species richness,

the relative size and the degree of specialization of the interacting species, the functional groups  
795 involved, and the magnitude of the climatic changes.

(6) To conclude, our review shows that although increased climatic variability is likely to have  
important biological consequences, this aspect of climate change has received little attention  
compared to changes in climatic averages. Given the prospect of increased climatic variability as a  
component of climate change in many locations, there is a clear need to increase research efforts to  
800 understand how this component of climate change influences physiological, ecological and  
evolutionary processes.

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## **IX. APPENDIX A1: USING TULJAPURKAR’S (1990) ‘SMALL-NOISE’ APPROXIMATION TO UNDERSTAND THE INFLUENCE OF INCREASED VARIANCE OF VITAL RATES ON THE STOCHASTIC POPULATION GROWTH RATE**

815 In this appendix, we explain how, in the case of independently and identically distributed (IID) environmental variation, the predictions about whether increased variance of a vital rate will increase or decrease the long-term stochastic population growth rate  $\lambda_s$  can be understood in terms of Tuljapurkar’s (1990) ‘small-noise’ approximation.

A population projection matrix integrates the effects of the vital rates into a single measure  
820 of the population growth rate. Much of the population theory that describes the effect of environmental variation on population growth relies on Tuljapurkar’s (1990) ‘small-noise’ approximation, derived using a stochastic projection matrix model and assuming the vital rate variances are not too large (hence ‘small noise’). In its simplest form, Tuljapurkar’s approximation relates the long-run stochastic population growth rate  $\lambda_s$  to the means, standard deviations, and  
825 correlations among vital rates as follows:

$$\log \lambda_s \approx \log \bar{\lambda} - \frac{1}{2\bar{\lambda}^2} \sum_{i,j} S_i S_j \sigma_i \sigma_j \rho_{i,j}.$$

[Note that this version of the approximation is written in terms of the underlying vital rates,  
830 accounting for the effects that each vital rate may have on multiple elements in the projection matrix; see Doak *et al.* (2005). For an alternative version based directly on the matrix elements, see Caswell (2001).] The means of the vital rates affect  $\bar{\lambda}$ , the asymptotic population growth rate

predicted by the average of all the annual projection matrices. The sum is taken over all the vital  
 rate pairs  $i, j$ . The sensitivity  $S_i$  is the slope of a graph of  $\bar{\lambda}$  versus vital rate  $i$  at that vital  
 835 rate's current value (mathematically, it is the partial derivative of  $\bar{\lambda}$  with respect to the vital rate,  
 evaluated at that rate's current value and keeping all other vital rates fixed). Sensitivity measures the  
 influence that each vital rate has on the population growth rate. The sensitivities appear in the  
 'small-noise' approximation because we must weigh the impact of variation in the vital rates by the  
 influence that each vital rate has upon annual population growth rate in order to predict the amount  
 840 of temporal variation in annual population growth, which is what affects the long-run growth rate  
 $\lambda_s$ . Temporal variation in low-sensitivity vital rates will produce less variation in the annual  
 population growth rate than will variation in high-sensitivity vital rates. The sensitivities are  
 computed directly from the mean projection matrix, just as  $\bar{\lambda}$  is, so they are influenced only by  
 the means of the vital rates. The parameters  $\sigma_i$  and  $\sigma_j$  are the standard deviations of the two  
 845 vital rates, and  $\rho_{ij}$  is the coefficient of correlation between vital rates  $i$  and  $j$ , positive values  
 indicating that the two rates tend to be relatively high in the same years (and relatively low in the  
 same years), and negative values meaning one tends to be low when the other is high. This simple  
 version of the 'small-noise' approximation assumes that there is no correlation in environmental  
 conditions across years (that is, we imagine choosing at random from a set of vital rate values each  
 850 year, without regard to whether we chose favourable or unfavourable rates in the previous year).

The 'small-noise' approximation becomes even simpler in the special case in which vital  
 rates vary independently of one another, so that the correlation  $\rho_{ij}$  is zero except when  $i = j$ , and  
 then it equals one. Thus, the term for vital rate  $i$  within the sum becomes simply  $S_i^2 \sigma_i^2$ , the  
 product of the squared sensitivity and the variance of the vital rate, which is always positive.  
 855 Because the entire sum is then positive, the minus sign in front of it means that an increase in the  
 variances of the vital rates will cause a *decrease* in the long-run population growth rate relative to  
 the rate ( $\bar{\lambda}$ ) that would occur in the absence of environmental variability. So, independent

environmental variation in the vital rates is detrimental to population growth. Furthermore, the greater is  $S_i$ , the sensitivity of vital rate  $i$ , the greater is the depressive effect on population growth of increasing its variance. This is the basis of the prediction that natural selection should favour life-history traits that reduce temporal variation in those vital rates that have the greatest influence on population growth, a prediction that has some empirical support (Pfister, 1998; Morris & Doak, 2004).

The picture is somewhat more complex when variation between vital rates is correlated, positively or negatively. If the sensitivities of two vital rates are both positive but the vital rates covary negatively, the term in the sum for that vital rate pair will be negative, which when multiplied by the minus sign outside the sum yields a positive term that *increases* the long-run population growth rate when the temporal variance (or standard deviation) of each vital rate increases. This positive contribution to long-run growth must be weighed against the  $S_i^2 \sigma_i^2$  terms in the sum for the two vital rates, which will still be positive, depressing growth. But taking both the positive and negative contributions in this case into account, it is possible for increasing variance in a vital rate with a relatively low sensitivity to cause the long-run growth rate to *increase*, by acting as a counterweight against variation in higher-sensitivity vital rates (Doak *et al.*, 2005). Another way that increasing vital rate variance can increase the long-run population growth rate is when a vital rate (such as one that represents shrinkage to a smaller size, or the rate of entering a dormant state) has a small negative sensitivity and a positive correlation with higher (but positive) sensitivity rates. Thus we cannot say categorically that increasing the variance of a demographic rate will invariably depress the population growth rate; it depends on the details of the life history (which influence the sensitivities) and on the way in which different vital rates co-vary.

880 **Table 1.** A sample of studies addressing phenotypic plasticity in plant functional traits in response to treatments of water or temperature. Experimental  
plants came from populations with contrasting temporal heterogeneity in soil moisture or temperature. In Outcome, + indicates greater plasticity in the  
population(s) from the more variable habitat(s), – indicates the opposite, and 0 refers to no differences in plasticity among populations. *N* pop =  
number of populations. Overall, there was a statistically significant trend for populations from more heterogeneous environments to show increased  
phenotypic plasticity ( $P < 0.05$ , sign-test).

885

Species ( <i>N</i> pop)	Environmental factor	Scale of temporal heterogeneity	Outcome	Reference
<i>Artemisia californica</i> (5)	Water	Between years	+	Pratt & Mooney (2013)
<i>Artemisia frigida</i> (2)	Water	Between years	+	Liu <i>et al.</i> (2014)
<i>Convolvulus arvensis</i> (2)	Water	Within year	+	Gianoli (2004)
<i>Convolvulus chilensis</i> (3)	Water	Between years	+	Gianoli & González-Teuber (2005)
<i>Leymus chinensis</i> (5)	Water	Between years	+	Liu <i>et al.</i> (2014)
<i>Plantago lanceolata</i> (29)	Temperature	Within year	–	Lacey <i>et al.</i> (2010)
<i>Polygonum persicaria</i> (2)	Water	Between years	0	Sultan & Bazzaz (1993)*
<i>Polygonum persicaria</i> (3)	Water	Within year	0	Heschel <i>et al.</i> (2004)
<i>Ranunculus flammula</i> (10)	Water	Between years	+	Cook & Johnson (1968)

Species ( <i>N</i> pop)	Environmental factor	Scale of temporal heterogeneity	Outcome	Reference
<i>Senna candolleana</i> (4)	Water	Between years	+	Lázaro-Nogal <i>et al.</i> (2015)
<i>Stipa grandis</i> (2)	Water	Between years	–	Liu <i>et al.</i> (2014)
<i>Stipa krylovii</i> (3)	Water	Between years	+	Liu <i>et al.</i> (2014)
<i>Taraxacum officinale</i> (2)	Water	Within year	+	Molina-Montenegro <i>et al.</i> (2010)
<i>Taraxacum officinale</i> (5)	Temperature	Within year	+	Molina-Montenegro & Naya (2012)

\* The authors did not compare plasticity levels between populations; the outcome was inferred after statistical comparison of *F*-ratios.

## FIGURE LEGENDS

**Fig. 1.** An example to illustrate the difference between changing the variance and the kurtosis (occurrence of extreme events) of a climatic variable. (A) Three contrasting distributions of a climatic variable are shown, one following the normal distribution with a variance of 1 (black), another following the normal distribution with a variance of  $5/3$  (blue), and the last following Student's  $t$  distribution with a variance of  $5/3$  (green). Thus, the black curve has lower variance than the blue curve but both have the same kurtosis, as by definition the normal distribution has zero kurtosis. In turn, the green curve has the same variance as the blue curve ( $5/3$ ) and is leptokurtic, with a kurtosis value of 6. Thus, it exhibits greater occurrence of extreme events than the other curves and has greater kurtosis than the black and blue curves. The three curves are centered on the same mean. (B) The same three curves shown in A with the ordinates in log scale, which helps when comparing the frequency of extreme events among the curves.

900

**Fig. 2.** Given the nonlinear relationships between performance and body temperature, mean performance differs between ectothermic organisms inhabiting constant and fluctuating environmental conditions. Coloured circles symbolize the performance of individuals kept at either a low (blue circle) or a high (red circle) constant temperature. Green circles denote performances of individuals kept at thermal cycles following a step function; arrows indicate the range of the step function. When the mean temperature lies below the thermal optimum ( $T_0$ ), thermal variation increases the mean performance. Contrarily, when the mean temperature exceeds the thermal optimum, thermal variation reduces the mean performance. Modified from Bozinovic *et al.* (2011a).

910 **Fig. 3.** The adaptive value of phenotypic plasticity under two different scenarios of climate change: (i) a temporal shift in environmental quality (and in its corresponding cue for the initiation of the

plastic response) (left column), and (ii) a temporal shift in environmental quality plus climatic variability (a short-term anomaly counteracting the climatic trend) (right column). (A, B) Seasonal variation in environmental quality in historical climate (black curve) *versus* climate change (red curve) scenarios. (A) Climate change = continuous trend. (B) Climate change = a shift in historical climate plus climatic variability. The level of environmental quality that acts as a cue for the initiation of the plastic response is indicated. (C, D) Reaction norms from three genotypes in the climate change scenarios depicted in A and B. Their responses to environmental cues (or lack thereof) are evidenced by a change in the slope of their reaction norms. It is assumed that the trait changes continually after the threshold is exceeded. The black genotype does not respond to the temporal shift in the appearance of the cue (C) or climatic variability (D), keeping the phenotypic trajectory corresponding to the historical climate. The red genotype has adjusted its reaction norm to the shift in the cue (C) or climatic variability (D). The purple genotype, together with adjusting its reaction norm as for the red genotype, shows increased plasticity (steeper slopes; C, D). The plateaus in the red and purple genotypes (D) arise when the environment goes back above the threshold in B. The orange dashed line indicates the occurrence of selection at the end of the season. (E, F) Adaptive landscape assuming directional selection, showing the relative position of the three genotypes in the scenarios of continuous climate change (E) and climatic variability (F), following the phenotypic trait expression observed at the time of selection (orange dashed line). (G, H) As for E and F, but this time assuming stabilizing selection when building the adaptive landscape.

**Fig. 4.** A partial summary of population theory describing the effect of a change in environmental variability on the long-term stochastic population growth rate,  $\lambda_s$ . Abbreviations: IID, independently and identically distributed; VR, vital rate; VRV, vital rate variance. References: 1: Tuljapurkar (1990); Pfister (1998); 2: Doak *et al.* (2005); 3: Morris *et al.* (2006); 4: Gross *et al.* (1998), Caswell & Kaye (2001), Tuljapurkar *et al.* (2003), Menges & Quintana-Ascencio (2004),

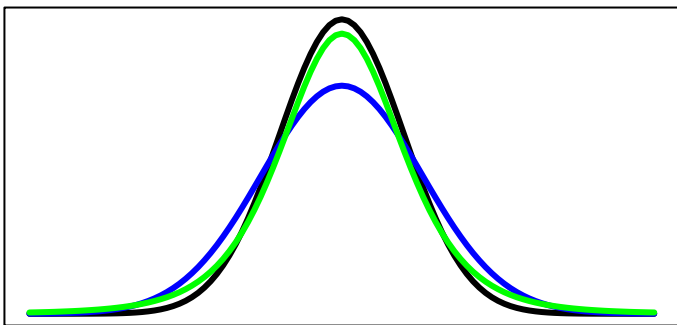
**Fig. 5.** In a changing climate, the mean and variance of environmental conditions may change in the same or in opposite directions, and may drive similar changes in the means and variances of vital rates.

**Fig. 6.** Mechanisms exacerbating phenological mismatches at geographic range limits. (A) Greater influence of harsh climatic events at range limits. In the example, the phenology of a consumer (blue) overlaps well with the average phenology of its resource (green). If the probability of occurrence of harsh climatic events that damage the resource is greater at range limits (black dotted line) than at the range center (grey dotted line), a phenological shift in the resource (red) should exacerbate the phenological mismatch at range limits. The black arrow indicates the direction of the shift in the plant phenology. (B) Decreased sampling probability of phenological tails at range limits. If the abundance of a key resource that is abundant throughout the geographic range (green) decreases towards range limits (red), statistical sampling will dictate a shorter resource phenology at range limits, with a later start and earlier end of the phenological period (dotted vertical lines). In the example, resource phenologies were generated by randomly sampling from a normal distribution, with the abundance away from range limits being five times greater than the abundance at range limits.



**A**

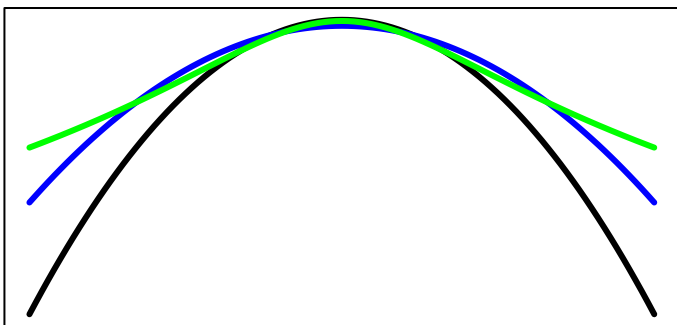
Probability of occurrence



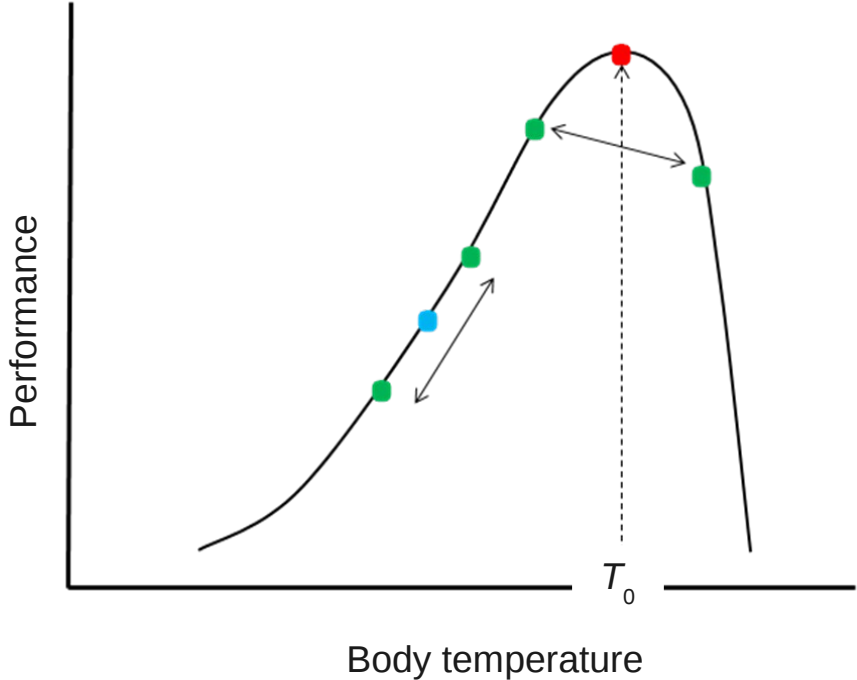
Climatic variable

**B**

Probability of occurrence

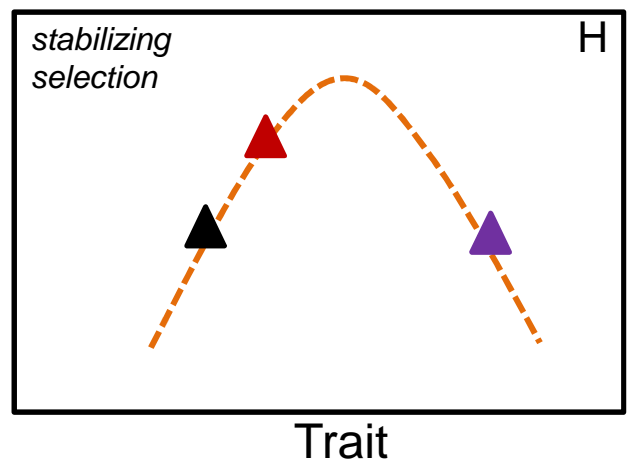
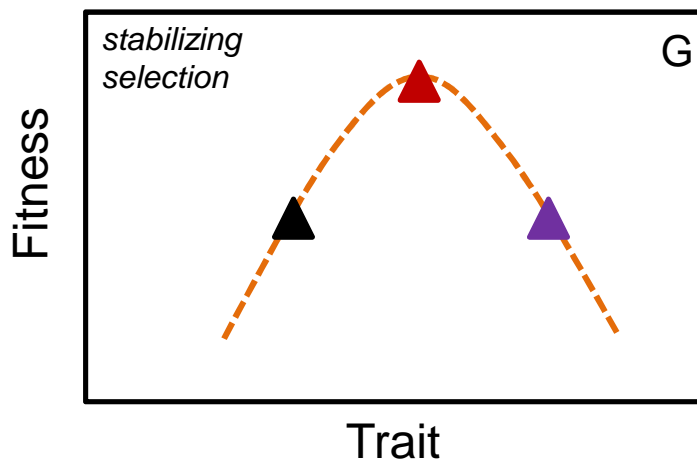
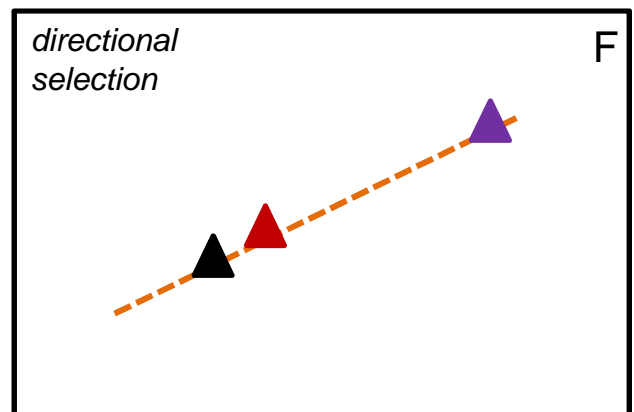
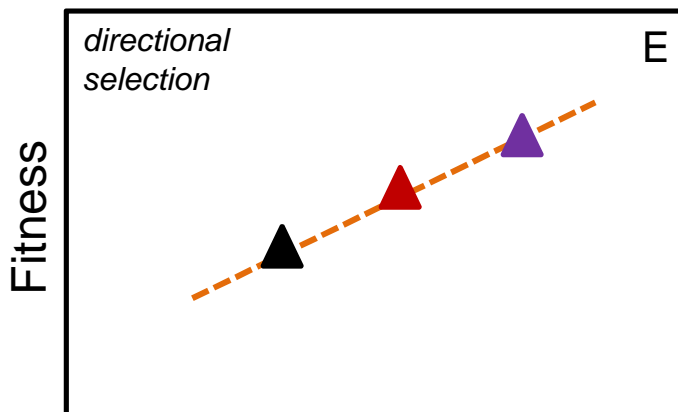
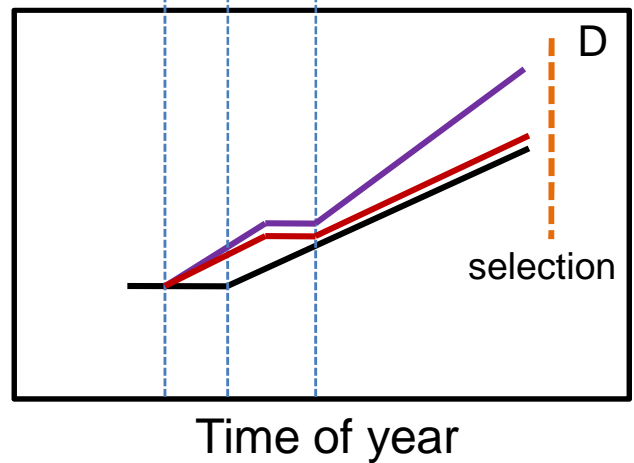
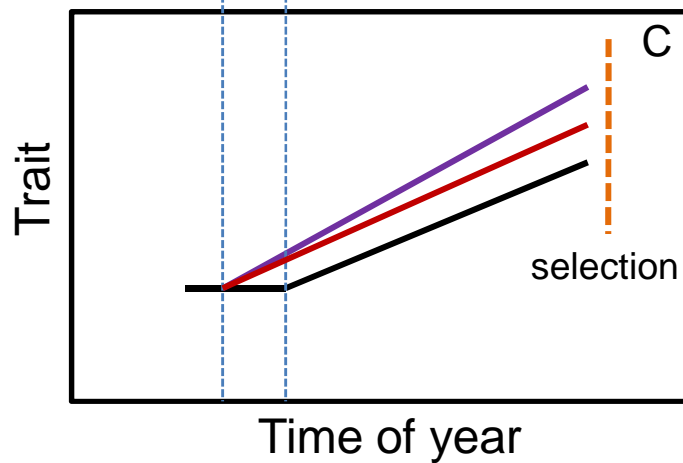
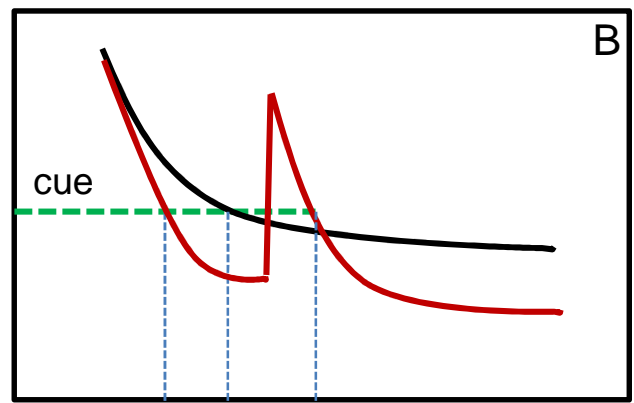
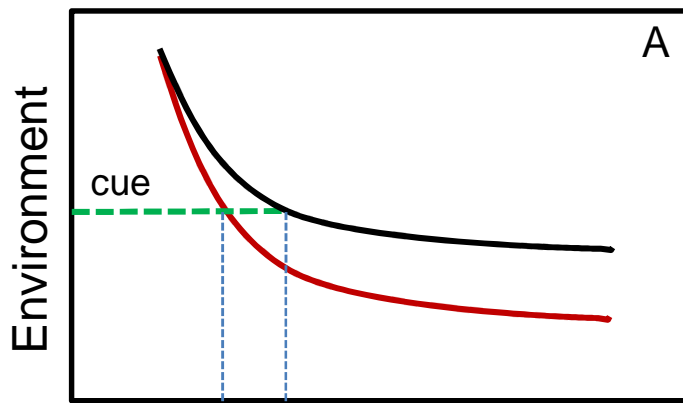


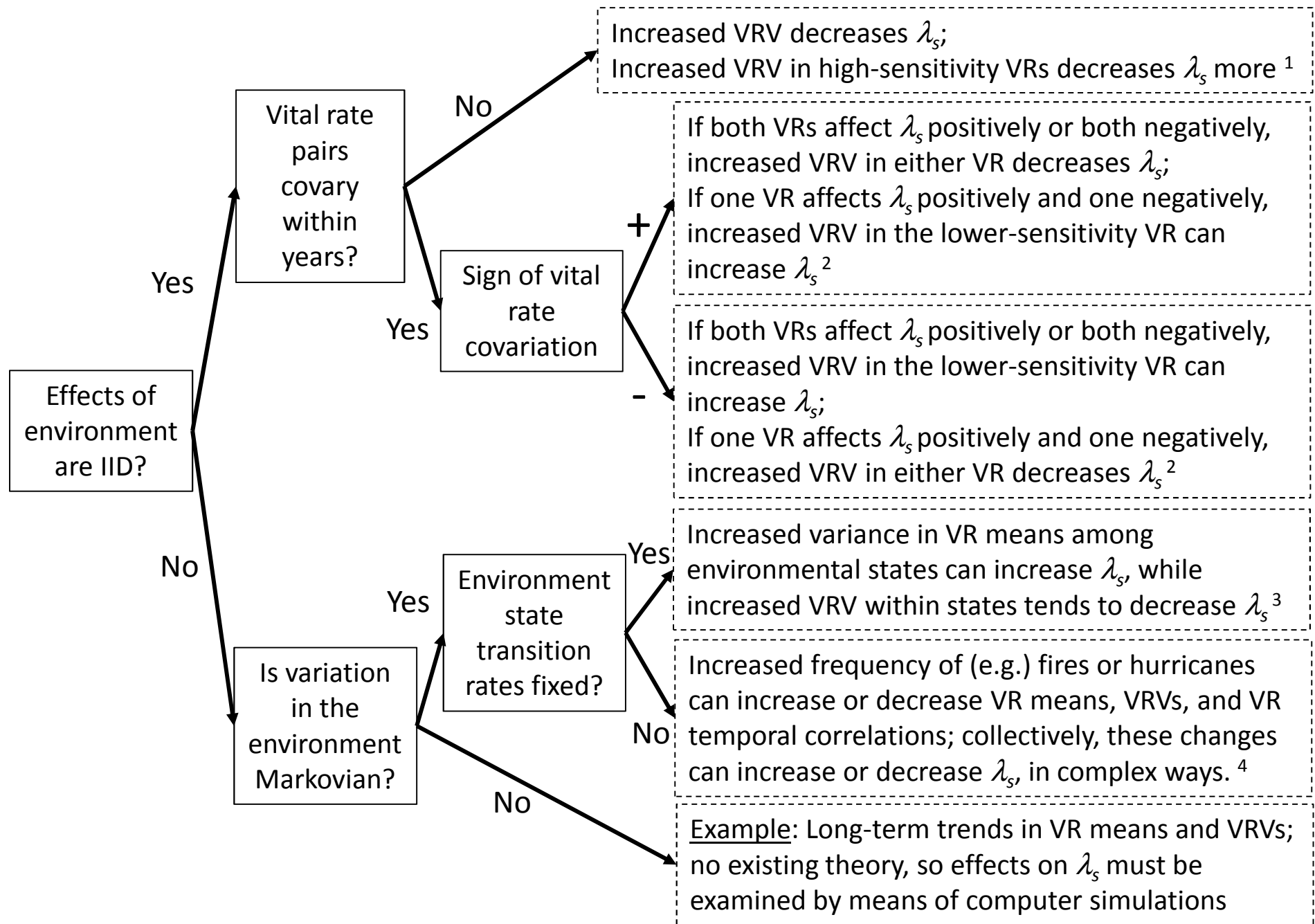
Climatic variable



Changes in climatic averages

Changes in climatic averages + variability

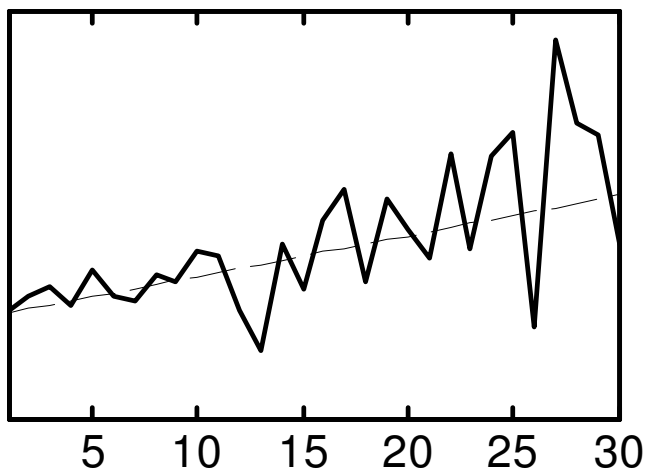




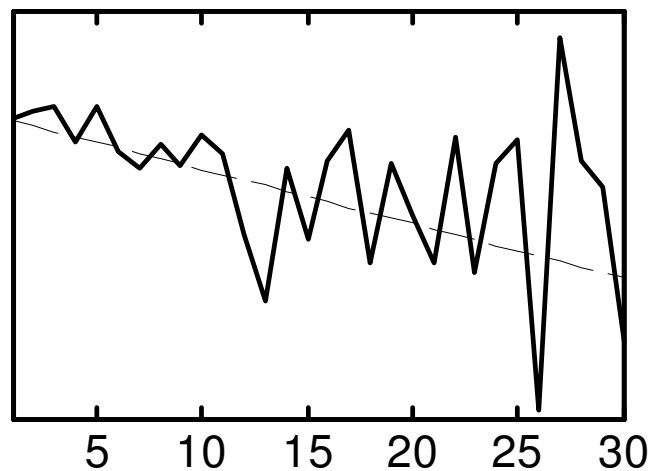
Variability  
increasing

Value of climate variable

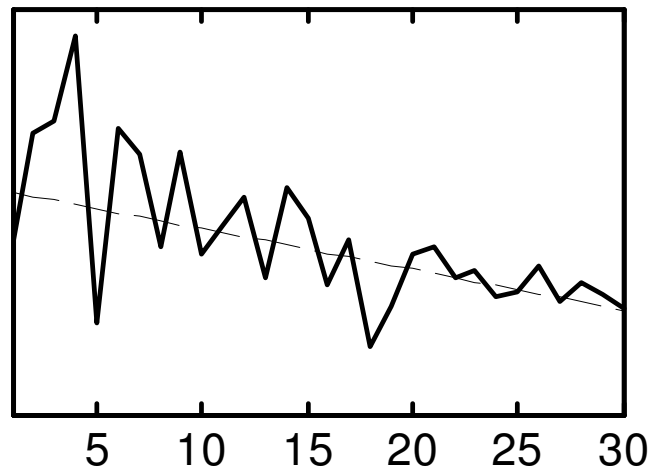
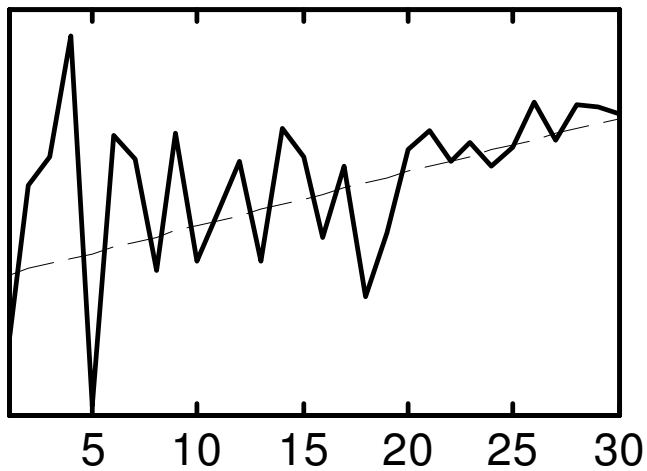
Mean increasing



Mean decreasing



Variability  
decreasing



Year

Resource or consumer abundance

